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Why do arms extract less oxygen than legs during exercise?


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Calbet, J. A. L., H.-C. Holmberg, H. Rosdahl, G. van Hall, M. Jensen-Urstad, and B. Saltin. Why do arms extract less oxygen than legs during exercise? Am J Physiol Regul Integr Comp Physiol 289: R1448–R1458, 2005. First published May 26, 2005; doi:10.1152/ajpregu.00824.2004.—To determine whether conditions for O2 utilization and O2 off-loading from the hemoglobin are different in exercising arms and legs, six cross-country skiers participated in this study. Femoral and subclavian vein blood flow and gases were determined during skiing on a treadmill at ~76% maximal O2 uptake (\(\dot{V}O_2\)max) and at \(\dot{V}O_2\)max, with different techniques; diagonal stride (combined arm and leg exercise), double poling (predominantly arm exercise), and leg skiing (predominantly leg exercise). The percentage of O2 extraction was always higher for the legs than for the arms. At maximal exercise (diagonal stride), the corresponding mean values were 93 and 85% (\(n = 3; P < 0.05\)). During exercise, mean arm O2 extraction correlated with the PO2 value that causes hemoglobin to be 50% saturated (P50: 0.93, P < 0.05), but for a given value of P50, O2 extraction was always higher in the legs than in the arms. Mean capillary muscle O2 conductance of the arm during double poling was 14.5 (SD 2.6) ml·min\(^{-1}\)·mmHg\(^{-1}\), and mean capillary PO2 was 47.7 (SD 2.6) mmHg. Corresponding values for the legs during maximal exercise were 48.3 (SD 13.0) ml·min\(^{-1}\)·mmHg\(^{-1}\) and 33.8 (SD 2.6) mmHg, respectively. Because conditions for O2 off-loading from the hemoglobin are similar in leg and arm muscles, the observed differences in maximal arm and leg O2 extraction should be attributed to other factors, such as a higher heterogeneity in blood flow distribution, shorter mean transit time, smaller diffusing area, and larger diffusing distance, in arms than in legs.

MUSCULAR OXYGEN UPTAKE depends on extrinsic factors such as O2 delivery and the intrinsic factors that regulate both the transfer of O2 from the erythrocytes to the mitochondria and the subsequent utilization of O2 in the mitochondria. However, the diffusive transfer of O2 is not only determined by intrinsic factors, because it also depends on mean capillary O2 tension. It is currently assumed that during exercise with a small muscle mass, intrinsic factors are the main determinants of peak local muscular \(\dot{V}O_2\), because the O2 delivery is extraordinary high (3, 44, 61). During exercise with a large muscle mass, the \(\dot{V}O_2\) peak of the lower extremities appears to be O2 delivery dependent (6, 7, 16, 33, 35, 57). O2 extraction across the lower extremities may reach maximal values between 90 and 92% of the arterial O2 content (CaO2), and the PO2 in the femoral vein may be close to 10 mmHg in active subjects (6, 7, 16), leaving little room for further extraction. However, in sedentary subjects, the maximal O2 extraction across the legs lies close to 70% of the CaO2, implying that their peak muscular \(\dot{V}O_2\) also may be limited by intrinsic factors (20). In physically active but non- arm-trained subjects, a low O2 extracting capacity has been reported for the arms (1, 11, 51, 70). Moreover, arm training resulted in only a marginal improvement in the O2 extraction of the arms (51). Therefore, the intrinsic factors may play an important role in limiting the maximal \(\dot{V}O_2\) attainable during maximal arm exercise. One of these roles could be differences in muscle capillarization that may affect both mean transit time (MTT) and diffusion conditions in the muscle. Moreover, primarily on the basis of experiments using the isolated hind-limb preparation in rats, it has been shown that mitochondrial oxidative capacity could determine the rate of O2 utilization and thereby O2 extraction (22, 37, 58). However, the limiting factor may be different in humans as suggested by the fact that prolonged bed rest decreases mitochondrial oxidative capacity and \(\dot{V}O_2\)max without reducing O2 extraction in humans (13, 62). Moreover, whole body \(\dot{V}O_2\) increases by 6% with hypoxia (fraction of inspired O2 = 0.5) during arm-cranking exercise (27). This observation is also compatible with a O2 delivery limitation of arm peak \(\dot{V}O_2\). However, the effect of hypoxia also could be explained, but only in part, by the increase in the amount of free O2 in the arterial blood in subjects without exercise-induced hypoxemia. Therefore, we hypothesize that O2 extraction across the arms is low because of a less efficient O2 off-loading from the hemoglobin compared with the legs.

The aim of this study was to determine, first, whether O2 extraction of the arm muscles is lower than that of leg muscles in humans with well-trained arm and leg muscles, at given systemic and regional absolute \(\dot{V}O_2\). Second, we assessed whether the conditions for the O2 off-loading from the hemoglobin are different for the arms and legs during submaximal and maximal combined arm and leg exercise in arm- and leg-trained humans. Third, we assessed whether differences in muscle oxidative capacity could account for the differences in O2 extraction between arm and leg muscles. To pursue these aims, we studied a group of well-trained cross-country skiers.

METHODS

Subjects. Six elite cross-country skiers, age 24 (SD 4) yr, height 180 (SD 6) cm, and weight 74 (SD 6) kg, volunteered to participate in the study. One week before the experiment, their maximal O2 uptake (\(\dot{V}O_2\)max) was 5.1 (SD 0.3) l/min or 72 (SD 4) ml·kg\(^{-1}\)·min\(^{-1}\), assessed during an incremental intensity test to exhaustion. The incremental exercise test was carried out on skiers using the diagonal
stride technique while skiing uphill with roller skis on a modified treadmill (Refax, Falun, Sweden). This \( \text{VO}_2 \max \) value is referred to as \( \text{VO}_2 \max \) DS. All subjects were informed about the possible risks and discomfort involved in the study before they gave their written consent to participate. This study was carried out according to the Declaration of Helsinki and was approved by the Ethical Committee of the Karolinska Institute, Stockholm, Sweden. The reason for choosing cross-country skiers was not only that they have well-trained arm and leg muscles but also that they are able to skillfully perform exercise with the upper or lower extremities, as well as exercise combining the upper and lower extremities in the upright position.

**Experiment preparation.** All subjects were familiar with the use of roller skis. On the experimental day, the subjects reported to the laboratory at 8:00 AM, and catheters were placed under local anesthesia (2% lidocaine) and advanced to the final position under fluoroscopic guidance, as previously described (9, 69). An 18-gauge catheter (Hydroch; Ohmeda, Swindon, UK) was inserted percutaneously, using the Seldinger technique, into the left or right femoral artery 2–5 cm below the inguinal ligament and was advanced 5–10 cm in the proximal direction. This catheter was connected to a blood pressure transducer positioned at the height of the fourth intercostal space (T100209A; Baxter, Unterschleissheim, Germany) and was also used to sample arterial blood. A 20-gauge catheter was inserted in the left femoral vein 2 cm below the inguinal ligament and was advanced 5–7 cm in the distal direction for femoral venous blood sampling. In the right femoral vein, a venous catheter with side holes (Radiopack TFE; Cook, Bjaerverskov, Denmark) was inserted and advanced ~5 cm proximal to the inguinal ligament for the injection of iced physiological saline solution. A thin polyethylene-coated thermostat (model 94-030-2.5F T.D. Edslab probe; Baxter, Irvine, CA) was inserted through the venous catheter for blood flow measurements by using the constant infusion thermodilution technique (3). An additional 18-gauge catheter also was inserted into the left femoral vein 2–3 cm below the inguinal ligament and was advanced under fluoroscopic guidance until the tip was positioned in the center of the right atrium, to sample blood from the right atrium. The last catheter, a Swan-Ganz triple-lumen catheter (model 132PS Edslab) was inserted into an ante-cubital vein and, under fluoroscopic guidance, was advanced into the subclavian vein until the tip was positioned at 5 cm before the merger with the jugular vein. One lumen was used for blood sampling and another for infusion of iced saline solution for blood flow measurements. Infuse temperatures were measured with a thermistor set in a flow-through chamber (model 93-505 Edslab) connected to the venous catheters. All sampling catheters were connected to a three-way stopcock and, along with the thermistor, were sutured to the skin to minimize the risk of movement during exercise.

A three-lead electrocardiogram (ECG) was displayed on a monitor during catheterization and the rest of the experimental procedures (Dialogue 2000, Danica, Copenhagen, Denmark). The ECG, blood pressure, and the temperatures registered by the thermistors, as well as the temperature at the thermistor, were recorded simultaneously with the data acquisition system (MacLab 16/s; ADInstruments, Sydney, Australia). Once the catheterization was finished, the subjects lay in the supine position for 180 min. One hour later, muscle biopsies were taken, and the blood was collected in ice-cold tubes that contained 10% O2 blood and was immediately centrifuged at 4°C for 10 min and stored at -80°C. Two hours after the merger with the jugular vein. One lumen was used for femoral vein temperature had stabilized at its new lower value. Blood flow was calculated on the basis of thermal balance principles, as detailed by Andersen and Saltin (3). Resting blood flow and arterial blood pressure were measured six times over 60 min and averaged. During submaximal exercise, blood flow measurements were performed in duplicate. The reported submaximal blood flow values represent the average of at least two measurements. At peak effort, the measurements were made within 1 min of exhaustion.

**Leg and Arm \( \text{VO}_2 \).** Leg and arm \( \text{VO}_2 \) values were computed separately using the Fick method, i.e., leg \( \text{VO}_2 = \text{leg blood flow} \times (\text{CaO}_2 - \text{CvO}_2) \), and arm \( \text{VO}_2 = \text{subclavian vein blood flow} \times (\text{CaO}_2 - \text{CvO}_2) \), where \( \text{CvO}_2 \) represents the \( \text{O}_2 \) content in the subclavian vein and \( \text{CvO}_2 \) represents the \( \text{O}_2 \) content in the subclavian vein.

**Histochémical and enzymatic analysis.** Serial transverse sections were stained for myofibrillar ATPase as described by Brooke and Kaiser (5). Muscle capillary density was analyzed, visualized, and quantified as described by Qu et al. (50). Muscle biopsies were analyzed for citrate synthase (CS) and 3-hydroxyacyl-CoA dehydrogenase (HAD) activity (69).

**Blood samples and analytical procedures.** Blood was sampled anaerobically in heparinized syringes and immediately analyzed for hemoglobin (Hb), \( \text{O}_2 \) saturation (OSBM hemoxymeter; Radiometer, Copenhagen, Denmark), blood pH, \( \text{CO}_2 \), and \( \text{O}_2 \) tension (ABL5; Radiometer). Blood gases were corrected for measured femoral vein blood temperature (femoral venous and arterial blood gases) and subclavian vein blood temperature (subclavian venous blood gases). Blood \( \text{O}_2 \) content was computed from the saturation (\( \text{SO}_2 \)) and Hb concentration (\( \text{Hb} \)), i.e., \((1.34 \times [\text{Hb}] \times \text{SO}_2) + (0.003 \times \text{PO}_2)\). Another blood sample was taken, and the blood was collected in ice-cold tubes that contained 10 µl of 33.3 M EDTA per milliliter of blood and was immediately centrifuged at 4°C for 10 min and stored at -50°C until analysis. Plasma was analyzed enzymatically for...
lactate (Roche Unikit; Hoffman-LaRoche, Basel, Germany) on an automatic analyzer (Cobas Fara; Roche Diagnostics, Basel, Switzerland). Before biochemical analysis, muscle biopsy samples were freeze-dried and dissected free of connective tissue, visible fat, and blood with the use of a stereomicroscope. The standard P50, defined as the value of PO2 that causes Hb to be saturated by 50% when the O2Hb equilibration curve is determined at 37°C, pH 7.40, PCO2 40 mmHg, was calculated from the whole set of arterial and venous gases obtained in each experiment. The in vivo P50 was calculated using Kelman’s equation (32). The in vivo P50 is the PO2 value that causes Hb to be saturated at 50% at the temperature, PCO2, and pH of the blood in the femoral and subclavian veins during exercise. Arm and leg muscle O2 conductance and mean capillary PO2 values were determined as previously described by Wagner (71, 73).

Capillary muscle O2 conductance and mean capillary PO2. To calculate the diffusing capacity for O2 (DO2), we used an iterative numerical integration procedure to find the value of O2 conductance (i.e., in ml·min⁻¹·mmHg⁻¹) that yields the measured femoral muscle venous PO2. The calculation of DO2 assumes 1) that the intracellular PO2 is negligibly small at VO2max (15, 54), 2) that the O2 remaining in the femoral and subclavian venous blood is wholly accountable for by diffusion limitation of O2 from the microcirculation to the mitochondria, and 3) that perfusion/VO2 heterogeneity and perfusional or diffusional shunt are negligible. Mean capillary PO2 is the numerical average of all computed PO2 values, equally spaced in time, along the capillary from the arterial to venous end (49).

Statistical analysis. Descriptive statistics were performed on each variable to confirm the assumptions of normality and homoscedasticity. The effect of the skiing technique during submaximal exercise on the dependent variables was assessed using a one-way repeated-measures analysis of variance (ANOVA). Mauchly’s test of sphericity was run before the ANOVA, and in case of violation of the sphericity assumption, the degrees of freedom were adjusted according to the Huynh and Feldt test. Pairwise comparisons were carried out with Tukey’s test. The relationship between muscular O2 conductance and O2 extraction was determined using linear regression. Maximal exercise values were obtained in only three subjects. Thus we decided not to perform comparisons between submaximal and maximal exercise. However, to test for differences between arm and leg variables during maximal exercise in these three subjects, we used a paired Student’s t-test. The significance level was set at P < 0.05. Data are expressed as means ± SD, unless otherwise stated.

RESULTS

O2 delivery and consumption. During submaximal exercise, whole body VO2 was similar during the different exercise conditions at 4 l/min, which represented ~76% of VO2max measured during diagonal skiing (VO2max DS; Table 1). At peak exercise, a VO2 close to 95% of VO2max DS was reached. The contribution of the legs to whole body VO2 varied between 46% (during double poling) and 67% (during leg skiing). Conversely, for the arms, the corresponding values were 37 and 9%, respectively (Table 1). VO2peak during double poling is a little lower (~86% of the VO2max DS) (Holmberg et al., unpublished observations). This means that compared with the specific double poling VO2peak, the skiers were working at 86% of the double poling VO2peak.

Activities of muscle oxidative enzymes. The activities of muscle oxidative enzymes have been previously reported (69) and are summarized in Table 4 together with some additional unpublished data also obtained from cross-country skiers.

O2 extraction. The effect of the exercise on the blood gases, pH, and lactate is depicted in Table 2. The systemic and leg VO2...
content arterial-venous differences (a-vDif) were lower during arm exercise compared with the other exercise modes, whereas the arm O₂ a-vDif was lowest during leg skiing (Fig. 2A). Regardless of the exercise mode, the percentage of O₂ extraction was always higher for the legs than for the arms, even during double poling (Fig. 2B). During diagonal skiing, the femoral vein O₂ extraction values were slightly greater when the exercise was preceded by the double poling bout than during the 40 min of continuous diagonal skiing (Fig. 2B).

The exercise PO₂ and O₂ content values were always lower in the femoral vein than in the subclavian vein, whereas the O₂ tension and content values in the blood of the right atrium were in between (Fig. 3, A and B). Leg O₂ extraction was closely related to arm O₂ extraction during submaximal exercise with the diagonal stride technique (r = 0.89, P < 0.05). As shown in Fig. 4A, leg mean O₂ extraction correlated closely with mean leg VO₂ across conditions (r = 0.89, P < 0.05; n = 6), whereas this relationship was not significant at the arm level (r = 0.61, P = 0.28; n = 6). During arm exercise, there also was a close relationship between the mean O₂ extraction and the corresponding P₅₀ value (r = 0.93, P < 0.05) (Fig. 4B).

O₂ off-loading and arm diffusing capacity. As shown in Table 3, during the 40 min of continuous diagonal skiing, the in vivo P₅₀ value was lower than during the second bout of diagonal exercise, which was performed just after the double poling (33.7 (SD 1.2) and 35.2 (SD 1.5) mmHg, respectively). During combined leg and arm exercise, the blood pH was similar in the femoral and subclavian veins (Fig. 3C). How-

Table 1. O₂ delivery and VO₂

<table>
<thead>
<tr>
<th></th>
<th>Resting</th>
<th>Diagonal Skiing</th>
<th>Double Poling</th>
<th>Diagonal Skiing</th>
<th>Leg Skiing</th>
<th>Diagonal Skiing (Maximal)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Systemic O₂ delivery</td>
<td>1.86 (SD 0.47)</td>
<td>5.22 (SD 0.59)</td>
<td>5.00 (SD 0.56)</td>
<td>4.95 (SD 0.49)</td>
<td>4.99 (SD 0.51)</td>
<td>5.70 (SD 0.95)</td>
</tr>
<tr>
<td>2-Leg O₂ delivery</td>
<td>0.33 (SD 0.12)</td>
<td>2.53 (SD 0.39)†</td>
<td>2.05 (SD 0.39)*</td>
<td>2.52 (SD 0.27)*†</td>
<td>3.00 (SD 0.24)</td>
<td>3.44 (SD 0.54)</td>
</tr>
<tr>
<td>2-Arm O₂ delivery</td>
<td>0.23 (SD 0.10)</td>
<td>1.10 (SD 0.49)*†</td>
<td>2.02 (SD 0.29)*</td>
<td>1.13 (SD 0.20)*†</td>
<td>0.68 (SD 0.10)</td>
<td>1.27 (SD 0.38)</td>
</tr>
<tr>
<td>Pulmonary VO₂</td>
<td>0.31 (SD 0.35)</td>
<td>3.94 (SD 0.32)</td>
<td>3.74 (SD 0.44)</td>
<td>4.00 (SD 0.29)</td>
<td>3.96 (SD 0.42)</td>
<td>5.07 (SD 0.69)</td>
</tr>
<tr>
<td>2-Leg VO₂</td>
<td>0.08 (SD 0.05)</td>
<td>2.19 (SD 0.37)*†</td>
<td>1.72 (SD 0.42)*</td>
<td>2.27 (SD 0.22)*†</td>
<td>2.64 (SD 0.27)</td>
<td>3.22 (SD 0.61)</td>
</tr>
<tr>
<td>2-Arm VO₂</td>
<td>0.06 (SD 0.02)</td>
<td>0.78 (SD 0.29)*†</td>
<td>1.38 (SD 0.22)*</td>
<td>0.84 (SD 0.12)*†</td>
<td>0.36 (SD 0.07)</td>
<td>1.09 (SD 0.36)</td>
</tr>
</tbody>
</table>

Values are means (SD) (in l/min) for O₂ delivery and O₂ uptake (VO₂). Maximal exercise values were obtained in only 3 subjects (not included in statistical analysis). *P < 0.05 compared with leg exercise. †P < 0.05 compared with double poling.

Table 2. Blood gases, pH, and lactate during skiing with different techniques

<table>
<thead>
<tr>
<th></th>
<th>Resting</th>
<th>Diagonal Skiing</th>
<th>Double Poling</th>
<th>Diagonal Skiing</th>
<th>Leg Skiing</th>
<th>Diagonal Skiing (Maximal)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PO₂, mmHg</td>
<td></td>
<td></td>
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<td></td>
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<tr>
<td>Femoral artery</td>
<td>101.4 (SD 2.9)</td>
<td>90.9 (SD 7.5)*‡</td>
<td>105.1 (SD 6.6)</td>
<td>95.1 (SD 6.9)*‡</td>
<td>93.6 (SD 6.9)*‡</td>
<td>87.0 (SD 6.1)</td>
</tr>
<tr>
<td>Femoral vein</td>
<td>39.7 (SD 7.1)</td>
<td>15.3 (SD 2.3)*‡</td>
<td>17.5 (SD 3.4)*</td>
<td>13.6 (SD 2.4)*‡</td>
<td>14.6 (SD 2.2)*‡</td>
<td>12.3 (SD 0.9)</td>
</tr>
<tr>
<td>Subclavian vein</td>
<td>37.9 (SD 7.6)</td>
<td>22.2 (SD 2.7)*‡</td>
<td>26.5 (SD 2.0)*</td>
<td>21.7 (SD 2.7)*‡</td>
<td>27.8 (SD 5.4)*</td>
<td>18.0 (SD 1.4)</td>
</tr>
<tr>
<td>Right atrium</td>
<td>42.7 (SD 1.7)</td>
<td>21.3 (SD 3.7)*‡</td>
<td>22.5 (SD 2.2)*</td>
<td>19.2 (SD 2.0)*</td>
<td>19.6 (SD 2.4)*‡</td>
<td>15.8 (SD 0.5)</td>
</tr>
<tr>
<td>P₅₀, mmHg</td>
<td></td>
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<tr>
<td>Femoral artery</td>
<td>196.2 (SD 4.2)</td>
<td>194.1 (SD 9.8)</td>
<td>192.7 (SD 9.6)</td>
<td>188.8 (SD 9.1)</td>
<td>192.4 (SD 7.3)</td>
<td>190.0 (SD 7.8)</td>
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<tr>
<td>Femoral vein</td>
<td>147.8 (SD 25.0)</td>
<td>26.2 (SD 7.3)*‡</td>
<td>32.3 (SD 12.5)*</td>
<td>18.4 (SD 7.8)*</td>
<td>23.4 (SD 7.8)*</td>
<td>12.5 (SD 4.2)</td>
</tr>
<tr>
<td>Subclavian vein</td>
<td>138.2 (SD 38.7)</td>
<td>54.6 (SD 13.0)*‡</td>
<td>61.3 (SD 17.4)*</td>
<td>47.5 (SD 15.7)*‡</td>
<td>89.4 (SD 22.3)*</td>
<td>27.9 (SD 5.2)</td>
</tr>
<tr>
<td>Right atrium</td>
<td>161.7 (SD 5.6)</td>
<td>47.4 (SD 12.0)*‡</td>
<td>48.4 (SD 8.3)*</td>
<td>36.0 (SD 10.8)*</td>
<td>39.5 (SD 10.0)*</td>
<td>20.1 (SD 3.1)</td>
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<tr>
<td>pH</td>
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<tr>
<td>Femoral artery</td>
<td>7.41 (SD 0.02)</td>
<td>7.36 (SD 0.02)*</td>
<td>7.34 (SD 0.02)*</td>
<td>7.32 (SD 0.05)*</td>
<td>7.35 (SD 0.02)*</td>
<td>7.33 (SD 0.03)</td>
</tr>
<tr>
<td>Femoral vein</td>
<td>7.38 (SD 0.02)</td>
<td>7.25 (SD 0.02)*</td>
<td>7.24 (SD 0.02)*</td>
<td>7.23 (SD 0.05)*</td>
<td>7.23 (SD 0.02)*</td>
<td>7.19 (SD 0.05)</td>
</tr>
<tr>
<td>Subclavian vein</td>
<td>7.39 (SD 0.02)</td>
<td>7.25 (SD 0.02)*</td>
<td>7.19 (SD 0.02)*</td>
<td>7.22 (SD 0.07)*‡</td>
<td>7.30 (SD 0.05)*</td>
<td>7.15 (SD 0.07)</td>
</tr>
<tr>
<td>Right atrium</td>
<td>7.39 (SD 0.02)</td>
<td>7.26 (SD 0.02)*</td>
<td>7.22 (SD 0.02)*</td>
<td>7.23 (SD 0.07)*‡</td>
<td>7.26 (SD 0.02)*</td>
<td>7.17 (SD 0.05)</td>
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<tr>
<td>Lactate level, mmol/l</td>
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<tr>
<td>Femoral artery</td>
<td>0.6 (SD 0.2)</td>
<td>2.6 (SD 1.0)*‡</td>
<td>7.5 (SD 1.7)*</td>
<td>6.3 (SD 3.4)*</td>
<td>5.0 (SD 2.0)*</td>
<td>8.1 (SD 1.9)</td>
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<td>Subclavian vein</td>
<td>0.7 (SD 0.2)</td>
<td>3.3 (SD 1.2)*‡</td>
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<td>6.4 (SD 4.7)*</td>
<td>4.7 (SD 2.0)*</td>
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<td>6.7 (SD 3.7)*</td>
<td>5.0 (SD 2.2)*</td>
<td>8.9 (SD 2.8)</td>
</tr>
</tbody>
</table>

Values are means (SD), SO₂, O₂ saturation. Maximal exercise values were obtained in only 3 subjects (not included in statistical analysis). *P < 0.05 compared with resting conditions. †P < 0.05 compared with leg exercise. ††P < 0.05 compared with double poling.
ever, during double poling, the blood pH was lower in the subclavian than in the femoral vein, whereas the opposite was true during leg skiing. The lower extraction capacity of the arm was not related to differences in the variables that regulate the unloading of O2 from the Hb, because the degree of acidification of blood, blood temperature (not shown), and PCO2 (Fig. 3D) were very similar in the effluent blood from the legs and arms. However, compared with the subclavian vein, a small, but significantly higher blood temperature (+0.6 degrees) and PCO2 (6–10 mmHg) were observed in the femoral vein during leg skiing. Thus the calculated in vivo P50 value was almost the same in the effluent blood from the arm muscles during double poling and the effluent blood from the leg muscles during leg skiing (35.9 (SD 1.7) and 35.8 (SD 1.5) mmHg, respectively, $P = 0.66$) (Table 3). Despite this high P50 value in the arms, which was significantly higher than the 34.6 (SD 1.5) mmHg observed in the legs, the O2 extraction was markedly higher in the legs than in the arms during double poling (Fig. 2B).

At maximal exercise with the diagonal technique, the arm and leg in vivo P50 values were 38.8 (SD 4.3) and 37.5 (SD 3.1) mmHg (mean of 3 subjects). In the latter condition, one skier was able to extract 97 and 89% of the O2 supplied to legs and the arms, respectively. This superb O2 extraction was achieved with in vivo P50 values of 41.0 and 43.7 mmHg, respectively.

The calculated mean capillary muscle O2 conductance of the arm muscles during double poling was 14.5 (SD 2.6) ml·min$^{-1}$·mmHg$^{-1}$, and the mean capillary PO2 was 47.7 (SD 2.6) mmHg. The corresponding values for the leg muscles during maximal exercise with the diagonal technique were 48.3 (SD 13.0) ml·min$^{-1}$·mmHg$^{-1}$ and 33.8 (SD 2.6) mmHg. Arm muscle capillary O2 conductance tended to correlate with maximal arm O2 extraction percentage ($r = 0.76$, $P = 0.08$).

### DISCUSSION

This study shows that in arms, but not in legs, mean O2 extraction is closely related to the mean in vivo P50 value. Moreover, for a given P50 value, the upper extremities extract less O2 than the lower extremities in humans with highly trained arm and leg muscles during exercise. This lower O2 extraction is associated with a lower O2 conductance in the upper compared with the lower extremities. Hence, for a given O2 demand, a greater O2 delivery is needed for exercising arm than leg muscles, which is the cause of the relatively high blood flow to the arms.

Because O2 extraction increases with exercise intensity, and the relative intensity during double poling was ~86% of double poling V˙O2_peak, O2 extraction should have been higher for the arms than for the legs. However, the experimental findings showed higher extraction in the legs than in the arms. This aspect confers additional robustness to our findings.

Arm O2 extraction in trained and untrained muscles. Rasmussen et al. (51) reported mean axillary vein O2 saturation values of 38% (extraction: 60%) during arm cranking at an intensity elicitng a heart rate of 170 beats/min. In the current investigation, the subclavian vein saturation during submaximal exercise achieved mean values between 24 and 31% (extraction: 68–75%). The high arm O2 extraction capacity in the current investigation is likely the result of several years of regular training, given that Rasmussen et al. (51) reported that after 5 wk of intensive arm training (1 h × 5 days/wk), the extraction capacity of the arm muscles was only slightly improved. During maximal exercise, the arm muscles of our skiers extracted 85% of the O2 supplied, which is higher than reported during maximal cycle ergometry in the leg muscles of untrained people before (59) and even after 9 wk of a training program resulting in a 35% higher V˙O2 max (59). These skiers attained in their arms the same O2 extracting values as we have observed in the legs of physically active subjects (V˙O2 max between 47 and 63 ml·kg$^{-1}$·min$^{-1}$) during upright maximal cycle ergometry (7, 68). Roca et al. (59) reported maximal leg extraction values of 72% in sedentary subjects that increased to 82% after 9 wk of training. Compared with these values, our skiers also reached remarkably high maximal O2 extraction values in their legs (93%). However, untrained
Subjects may reach rather high leg O₂ extractions after bed rest (62) or during ischemic exercise (46).

Several factors may account for the observed differences in muscular O₂ extraction, which depends on the interaction of the following: 1) kinetics of O₂ off-loading from the Hb; 2) capillary muscle O₂ conductance (72); 3) blood flow, mean transit time (47), and degree of mismatch between the metabolic demand and blood flow distribution and/or degree of shunt (48); and 4) muscle maximal oxidative capacity (22) and exercise intensity.

Kinetics of O₂ off-loading from Hb. The off-loading of O₂ from Hb during exercise is facilitated by acidification of the
blood and the increase of temperature and \( P_{CO_2} \) (67). Experiments in dogs have shown that \( O_2 \) extraction at \( V_{O_2_{max}} \) decreases when the \( P_{50} \) of the Hb is reduced from its normal value of 32 to 23 mmHg (24), whereas it increases when the \( P_{50} \) is raised to 53.2 mmHg (56). In agreement, we observed a close correlation between \( O_2 \) extraction and \( P_{50} \). Moreover, during diagonal skiing, slightly higher femoral vein \( O_2 \) extraction values were observed after than before the double poling bout, which elicited a high degree of blood acidification, despite similar \( V_{O_2} \) and leg blood flows in both phases of diagonal skiing. The latter implies that for a given \( V_{O_2} \), facilitating the off-loading of \( O_2 \) from the Hb is associated with increased arm and leg \( O_2 \) extraction values during exercise. Thus our experiment shows that the conditions for the off-loading of \( O_2 \) from the Hb may influence \( O_2 \) extraction in the arm and leg muscles. However, despite small differences in blood temperature and the greater release of lactate and protons during the arm exercise (29, 69), the conditions at which the \( O_2 \) dissociation curve of Hb operates are similar in the arm and leg muscles (see Fig. 3). The lower arm \( O_2 \) extraction capacity cannot be explained by a slower off-loading of \( O_2 \) from the Hb in the arms, because the in vivo affinity of Hb for \( O_2 \) was similar in the arms and legs during both submaximal and maximal exercise.

**Capillary muscle \( P_{50} \) and \( O_2 \) conductance.** In this investigation we calculated the capillary muscle \( O_2 \) conductance by assuming that mitochondrial \( P_{50} \) is very close to 0–2 mmHg (47). The MTT of the erythrocytes crossing the muscle capillaries during maximal exercise is estimated as \( MTT = CBV/MBF \), where CBV is the capillary blood volume and MBF is the muscle blood flow. In turn, the capillary volume may be calculated from the capillary density (31). We did not measure the capillary density in the current investigation, but from previous studies in elite cross-country skiers of the same level of performance as the present subjects, it appears that the capillary density is similar in arm and leg muscles. For example, if at maximal exercise the active muscle mass is 4 and 11 kg in the arm and legs, respectively, the calculated MTT values will be 672 and 674 ms, respectively (31). However, the corresponding values will be 1,195 and 1,198 ms if an inner mean capillary diameter of 6.0 \( \mu \)m rather than 4.5 \( \mu \)m is assumed. What these numbers suggest is that MTT is rather high in both upper and lower extremities at maximal exercise. If MTT does not appear to explain the observed differences in \( O_2 \) extraction between the arms and legs, however, heterogeneity in macrovascular MTT has been related to \( O_2 \) extraction during low exercise intensity in humans (30). Nevertheless, macrovascular MTT heterogeneity decreases with exercise intensity, but the degree of skeletal muscle capillary MTT heterogeneity existing at maximal exercise remains unknown. In theory, MTT values lying between 0.3 and a little more than 1 s are possible (63).

In our experimental conditions, where exercise is performed at maximal intensity, there is no reason to suspect a mismatch, shunt, or differences in capillary length between the perfused and the active fibers (72) in arm and leg muscles (14, 26). However, we cannot rule out a greater degree of heterogeneity in the distribution of blood flow between muscles in the arms.
than in the legs. In fact, electromyographic activity data show large interindividual differences in the degree of recruitment of upper arm muscles during double poling (25). Lower O₂ extraction may be expected in the muscles that are less activated, hence also contributing to a low mean O₂ extraction of the whole extremity. Moreover, upper compared with lower extremities differ in degree of freedom of movement. Leg flexion and extension is performed in a rigid back and forward pattern with only minor variation in the activation of the different muscles and their functional portions. Arm and shoulder muscles, on the other hand, perform their movements in different positions, resulting in larger variation in the degree of muscle recruitment. Hence the mass of active muscles in the arm and shoulder region may be overestimated in cross-country skiing. In turn, the actual MTT across the active arm and shoulder region may be overestimated in cross-country skiing. In fact, endurance training induces an increase in the number of capillaries around each fiber (i.e., the amount of capillary-to-muscle fiber exchange surface) is slightly higher in the leg than in the arm muscles because of the higher mean fiber cross-sectional area of the arm muscles. Using the data in Table 4, we have calculated a mean capillary index of 0.011 and 0.010 capillaries per micrometer of capillary muscle O₂ conductance and the activities of CS and HAD. Previous studies also have reported lower CS and HAD activities in the triceps brachii compared with the vastus lateralis muscles of our skiers (69). However, no relationship was observed between capillary muscle O₂ conductance and the activities of CS and HAD. The transfer of O₂ from the Hb to the mitochondria should occur their maximal oxidative capacity even when exercising at VO₂ max with a small muscle mass (27). In fact, it has been estimated that only 20% of the muscle mass has to be recruited to tax the maximal oxidative capacity in humans (4). It is difficult to establish to what extent the role of this reduced oxidative capacity may be in the lower O₂ extraction capacity of the arms, but some animal in vitro experiments support this possibility (22, 37, 58) while others do not (36). In addition, the fact that VO₂ max increases with hyperoxia (29) and 30% reduction in cytochrome oxidase and of CS activity, respectively (36). Despite the fact that the arm oxidative capacity of the elite cross-country skiers is lower than their leg muscles (39), it is likely far in excess of their respective maximal O₂ demand (4). In support, we can argue

### Table 4. VO₂ max, capillary density, CS activity, HAD activity, % ST, and capillary index of cross-country skiers

<table>
<thead>
<tr>
<th>Muscle</th>
<th>VO₂ max, mL·kg⁻¹·min⁻¹</th>
<th>Capillary Density, cap/mm²</th>
<th>CS Activity, μmol·g⁻¹·dry wt⁻¹·min⁻¹</th>
<th>HAD Activity, μmol·g⁻¹·dry wt⁻¹·min⁻¹</th>
<th>%ST</th>
<th>Mean Fiber Area, μm²</th>
<th>Cap/Fiber</th>
<th>Capillary Index</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vastus lateralis</td>
<td>76</td>
<td>3⁰</td>
<td>654</td>
<td>88</td>
<td>74</td>
<td>72</td>
<td>5,200</td>
<td>3.4</td>
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<tr>
<td>Vastus lateralis</td>
<td>76</td>
<td>4⁰</td>
<td>553</td>
<td>82</td>
<td>66</td>
<td>61</td>
<td>5,600</td>
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<td>0.0116</td>
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<tr>
<td>Vastus lateralis</td>
<td>72</td>
<td>6⁰</td>
<td>51</td>
<td>38</td>
<td>38</td>
<td>38</td>
<td>5,400</td>
<td>3.0</td>
<td>0.0115</td>
</tr>
<tr>
<td>Deltoïd (posterior)</td>
<td>76</td>
<td>3⁰</td>
<td>582</td>
<td>76</td>
<td>61</td>
<td>61</td>
<td>5,300</td>
<td>3.1</td>
<td>0.0118</td>
</tr>
<tr>
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<td>76</td>
<td>4⁰</td>
<td>560</td>
<td>68</td>
<td>60</td>
<td>60</td>
<td>5,400</td>
<td>3.0</td>
<td>0.0115</td>
</tr>
<tr>
<td>Gastrocnemius (lateral)</td>
<td>72</td>
<td>3⁰</td>
<td>38</td>
<td>22</td>
<td>22</td>
<td>22</td>
<td>5,300</td>
<td>3.1</td>
<td>0.0110</td>
</tr>
<tr>
<td>Triceps (lateral)</td>
<td>72</td>
<td>8⁰</td>
<td>492</td>
<td>59</td>
<td>59</td>
<td>59</td>
<td>4,800</td>
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<tr>
<td>Triceps (lateral)</td>
<td>71</td>
<td>9⁰</td>
<td>415</td>
<td>55</td>
<td>43</td>
<td>43</td>
<td>6,400</td>
<td>3.1</td>
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<tr>
<td>Vastus lateralis</td>
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<td>10</td>
<td>422</td>
<td>49</td>
<td>31</td>
<td>31</td>
<td>6,100</td>
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<tr>
<td>Triceps (lateral)</td>
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<td>5³</td>
<td>373</td>
<td>39</td>
<td>20</td>
<td>20</td>
<td>7,400</td>
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<tr>
<td>Rectus femoris</td>
<td>17³</td>
<td>411</td>
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<td></td>
<td></td>
<td></td>
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<td></td>
<td>0.0008</td>
</tr>
<tr>
<td>Triceps (lateral)</td>
<td>17³</td>
<td>536</td>
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<td></td>
<td></td>
<td></td>
<td>4.9</td>
<td></td>
<td>0.0089</td>
</tr>
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</table>

VO₂ max, maximum O₂ uptake; CS, citrate synthase; HAD, 3-hydroxyacyl-CoA dehydrogenase; %ST, percentage of slow-twitch fibers; Cap, no. of capillaries. Capillary index is the mean no. of capillaries around each fiber per mean muscle fiber perimeter; n represents the no. of cross-country skiers: *top-level skiers, close to elite-level skiers, +women and 10 men skiers (VO₂ max not reported). Unpublished data.
that rowing $V_{O2max}$ increases with hyperoxia in elite rowers, a discipline in which there is a substantial contribution of the arms to the exercise $V_{O2max}$ (41, 45). In addition, Clausen et al. (11) showed that after 5 wk of leg training, the whole body $V_{O2}$ during maximal arm cranking increased by 10%, whereas the arm arterial-venous difference in $O_2$ content remained unchanged. Thus our data and those summarized in Table 4 suggest that the small difference in oxidative capacity between arm and leg muscles has a minor role, if any, in the lower maximal arm $O_2$ extraction capacity.

*Increased arm vascular reactivity?* Under resting conditions, the forearm has higher vasodilatory responsiveness to acetylcholine, substance P, nitroprusside (40), and isoprotanol (28) and lower vasoconstricting responsiveness to phenylpropanolamine ($α_1$ agonist) (43). These functional differences between arms and legs suggest increased ability to maintain vasodilation in the arms than in the legs under some level of ongoing sympathetic activation. Thus our data could be interpreted to indicate that the lower $O_2$ extraction in the arms only reflects a compensation for the relatively greater perfusion of the arms compared with the legs. However, if the reason for the lower $O_2$ extraction of the arms was that the arms vasodilated in excess, then $O_2$ extraction should have been similar in arms and legs at peak exercise with the diagonal style, because during this condition, perfusion of the arm and legs is limited by the pumping capacity of the heart (9). Nevertheless, peak arm blood flow reached a much lower value during maximal combined arm and leg exercise than during double poling, while $O_2$ extraction remained at a lower level in the arms than in the legs. Although the reported higher vascular reactivity of the forearm at rest may facilitate the vasodilatory response in the arms during combined arm and leg exercise, it remains unknown whether vascular reactivity remains higher in the arm than in the leg muscles during exercise. It may be that it does not, given that vascular reactivity during exercise is modulated by the compound action of several vasodilating and vasoconstricting agents (12). For example, despite a similar sympathetic activation in arms and legs in response to the cold pressor test, vascular resistance increases in the arms but not in the legs (28). Thus it is possible that during near-maximal combined arm and leg exercise, the exercise-induced elevation of muscle sympathetic nerve activity is likely more efficiently counteracted in the legs than in the arms, i.e., reducing the vasodilatory response of the arms more than that of the legs (66). This mechanism will give the perfusing priority to the main locomotory muscles. However, in case of competition for blood flow between the leg muscles and the respiratory muscles, the perfusion priority is likely given to the respiratory muscles (19).

*Potential limitations.* Although limited by the fact that $O_2$ extraction at maximal exercise intensity was not determined during isolated exercise with either the upper and the lower extremities, our results are clear: $O_2$ extraction across the upper extremities is lower than oxygen extraction across the lower extremities at a given absolute and relative exercise intensity. Given the fact that venous effluent blood from either extremity drains blood principally coming from the active muscles, the skin, and the bone marrow, the calculated extraction values represent the mean extraction capacity of the whole extremity. In theory, the skeletal muscles of both extremities could have a similar $O_2$ extraction capacity if the degree of "venous admixture," i.e., the amount of blood coming from the bone marrow and the cutaneous circulation at the sampling point, is higher in the arms than in the legs. However, the magnitude of the differences in fractional $O_2$ extraction between legs and arms is too high to be accounted for by different degrees of venous admixture. Of all potential contributors to venous admixture, skin blood flow is quantitatively the most important. Skin blood flow increases with body temperature. In these experiments we took measurements of venous $P_{O2}$ and $O_2$ extraction after 12 min of diagonal skiing, and they were repeated after 24 and 36 min of diagonal skiing (data not shown). During this period, exercise intensity was constant, but body temperature increased significantly from 38.1 to 38.6°C. Despite this increase in body temperature, and likely in skin blood flow (17), no increase in venous $P_{O2}$ or reduction in $O_2$ extraction was observed in arms or legs. This suggests that the degree of admixture is small and is not accentuated by a condition that increases skin blood flow. Moreover, we have calculated that during double poling, venous admixture in the upper extremities should have been at least 1.1 l/min greater than in the lower extremities in case the skeletal muscle $O_2$ extraction values in arm muscles during double poling were similar to the $O_2$ extraction in the leg muscles during leg skiing. This means that total venous admixture should have been 2.2 l/min higher in the upper than in the lower extremities, which does not seem reasonable because it would imply an impossible regional difference in cutaneous vasodilation between the upper and the lower extremities (34). Moreover, forearm exercise experiments were performed in the 1960s with venous catheters placed in deep forearm veins (under X-ray control) to avoid skin contamination and with a cuff inflated around the wrist at >200 mmHg to exclude the circulation of the hand (74). In these experiments, maximal exercise extraction values were lower than reported in the current investigation for the whole upper extremity (74).

Even with our state-of-the-art technology it is impossible to know the accurate amount of muscle mass activated during arm or leg exercise. Even worse, it is not possible to know to what extent the motor units are activated. The latter limitation also applies to human models of localized exercise such as the leg extension model. This problem holds for both extremities. The only way to circumvent this limitation is by using animal preparations, which may be stimulated electrically to activate maximally all motor units. However, these preparations show lower $D_O2$ values than reported in vivo simply because with these preparations $V_{O2peak}$ values in most models are ~50% of the real in vivo $V_{O2peak}$ values. Our lower $D_O2$ values agree well with the observation of lower $O_2$ extraction capacity for the arms. For an active muscle mass of 5 and 11 kg in arms and legs, a local muscle $V_{O2}$ of 138 and 147 ml/kg muscle mass can be calculated in the arms during double poling and in the legs during maximal diagonal stride, respectively. The corresponding "muscle-mass-normalized $D_O2$" is 2.9 and 4.4 ml·min⁻¹·mmHg⁻¹·kg muscle mass⁻¹. Arm muscle mass-normalized $D_O2$ would have matched leg muscle mass-normalized $D_O2$ for an active muscle mass of 3.3 kg in the arms and 11 kg in the legs. This proportion 3.3/11 (arm/leg active muscle mass) allows for a match in muscle mass-normalized $D_O2$ between arm and leg muscles but is simply impossible, because the $V_{O2peak}$ per kilogram of active muscle mass would have been 209 and 147 ml/kg, i.e., 42% higher in the arm than in leg.
ARM AND LEG MAXIMAL MUSCLE O₂ EXTRACTION


